

Physical limits to biochemical signaling

William Bialek* and Sima Setayeshgar†

Joseph Henry Laboratories of Physics and Lewis–Sigler Institute for Integrative Genomics, Princeton University, Princeton, NJ 08544

Communicated by Curtis G. Callan, Jr., Princeton University, Princeton, NJ, May 25, 2005 (received for review February 2, 2005)

Many crucial biological processes operate with surprisingly small numbers of molecules, and there is renewed interest in analyzing the impact of noise associated with these small numbers. Twenty-five years ago, Berg and Purcell showed that bacterial chemotaxis, where a single-celled organism must respond to small changes in concentration of chemicals outside the cell, is limited directly by molecule counting noise and that aspects of the bacteria's behavioral and computational strategies must be chosen to minimize the effects of this noise. Here, we revisit and generalize their arguments to estimate the physical limits to signaling processes within the cell and argue that recent experiments are consistent with performance approaching these limits.

A striking fact about biological systems is that single molecular events can have macroscopic consequences. The most famous example is, of course, the storage of genetic information in a single molecule of DNA, so that changes in the structure of this single molecule (mutations) can have effects on animal behavior and body plan from generation to generation (1). But the dynamics of individual molecular interactions can influence behavior on much shorter time scales. Thus, we (and other animals) can see when a single molecule of rhodopsin absorbs a photon (2), and some animals can smell a single molecule of airborne odorant (3). Even if a single molecular event does not generate a specific behavior, the reliability of behavior still can be limited by inevitable fluctuations associated with counting random molecular events. Thus, the visual system has a regime where perception is limited by photon shot noise (4, 5), and the reliability with which bacteria can swim up a chemical gradient appears to be limited by noise in the measurement of the gradient itself (6). It is an open question whether biochemical signaling systems within cells operate close to the corresponding counting noise limits.

The analysis of bacterial chemotaxis by Berg and Purcell (6) provided a clear intuitive picture of the noise in “measuring” chemical concentrations. Their argument was that if we have a sensor with linear dimensions a , we expect to count an average of $\bar{N} \sim \bar{c}a^3$ molecules when the mean concentration is \bar{c} . Each such measurement, however, is associated with a noise $\delta N_1 \sim \sqrt{\bar{N}}$. A volume with linear dimension a can be cleared by diffusion in a time $\tau_D \sim a^2/D$, so if we are willing to integrate over a time τ we can make $N_{\text{meas}} \sim \tau/\tau_D$ independent measurements, reducing the noise in our estimate of N by a factor of $\sqrt{N_{\text{meas}}}$. The result is that our fractional accuracy in measuring N , and hence in measuring the concentration c itself, is given by

$$\frac{\delta c}{\bar{c}} = \frac{\delta N}{\bar{N}} = \frac{1}{\sqrt{\bar{N} N_{\text{meas}}}} = \frac{1}{\sqrt{Da\bar{c}\tau}}. \quad [1]$$

A crucial claim of Berg and Purcell (6) is that this result applies when the sensor is a single receptor molecule, so that a is of molecular dimensions, as well as when the sensor is the whole cell, so that $a \sim 1 \mu\text{m}$.

The discussion by Berg and Purcell (6) made use of several special assumptions that we suspect are not required, which leads to some clear questions: For interactions of a substrate with a single receptor, does Eq. 1 provide a general limit to sensitivity, independent of molecular and biochemical details? Can we understand explicitly how correlations among nearby

receptors result in a limit like Eq. 1 but with a reflecting the size of the receptor cluster? Do the spatial correlations among nearby receptors have an analog in the time domain, so that there is a minimum averaging time required for noise reduction to be effective? Finally, if we can establish Eq. 1 or its generalizations as a real limit on sensitivity for any signaling process (Fig. 1), we would like to know if cells actually operate near this limit.

We address these questions within the general framework of statistical mechanics through analysis of intrinsic fluctuations of the receptor–ligand system. In most cases that we know about, biochemical signaling molecules are thought to interact with their receptors through some kinetic processes that lead to equilibrium between bound and unbound states. In this case, fluctuations in occupancy of a binding site are a form of thermal noise. Rather than tracing through the consequences of different microscopic hypotheses about the nature of the interaction between signaling molecules and their targets, this connection to thermal noise allows us to use the fluctuation–dissipation theorem (7–9), which relates noise levels to macroscopic kinetics in the same way that Einstein connected the statistics of Brownian motion to the macroscopic frictional forces on the Brownian particle.

Our main result is a derivation of the accuracy limit to which biochemical receptors are able to measure concentrations of signaling molecules. Although analysis of fluctuations in chemically reacting systems has been an active area in theoretical chemistry (10–14), most existing approaches are based on the Fokker–Planck or Langevin equations and have focused on the detailed connection between kinetic parameters and observable fluctuation spectra. In contrast, our goal is to establish, if possible, general limits on the sensitivity or accuracy of signaling systems that are independent of the often unknown kinetic details. We begin with a simple example of binding to a single receptor to present the fluctuation–dissipation theorem for chemical kinetic systems and to show that we can recover conventional results. By considering the coupled binding and ligand diffusion processes, we derive the accuracy limit to measuring the concentration of a diffusing ligand, where we find a contribution from the chemical kinetics of the measurement process as well as a lower bound that depends only on molecule counting noise. Within the same framework, we extend this result from a single receptor to multiple, noninteracting receptors, addressing the more complex case of cooperative interactions among receptors elsewhere (W.B. and S.S., unpublished results). Our results can be summarized by saying that the intuitive estimates of Berg and Purcell in fact correspond to a “noise floor” that is independent of kinetic details; real systems can be noisier but not more precise than this limit.

We compare our results with two recent quantitative experiments on intracellular signaling in *Escherichia coli*, regulation of gene expression by transcription factors and control of the flagellar motor by the “response regulator” CheY~P, and find

*To whom correspondence should be addressed. E-mail: wbialek@princeton.edu.

†Present address: Department of Physics, Indiana University, Bloomington, IN 47405.

© 2005 by The National Academy of Sciences of the USA

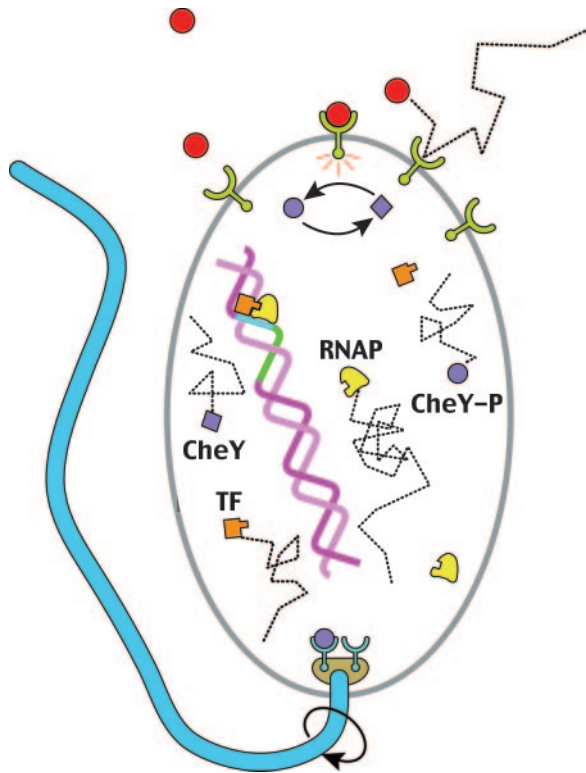


Fig. 1. Measuring the concentration of a signaling molecule by a biological sensor, which in turn controls downstream events, is a generic task. Here, several examples are depicted schematically for *E. coli*. Binding of attractant/repellent molecules to a surface receptor complex modulates the rate of autophosphorylation of the associated kinase. This change in kinase activity results in a corresponding concentration change of the internal signaling molecule, CheY~P, that controls the direction of flagellar motor rotation. Also shown is transcription initiation, where the promoter region can be regarded as a sensor for transcription factors (TF). These proteins, whose concentrations vary depending on the cell cycle and external cues, determine whether or not RNA polymerase (RNAP) turns on a gene.

that the performance of the cell is near the limit set by diffusive counting noise.

Theory

Binding to a Single Receptor. Consider a binding site for signaling molecules, and let the fractional occupancy of the site be n . If we do not worry about the discreteness of this one site, or about the fluctuations in concentration c of the signaling molecule, we can write a kinetic equation

$$\frac{dn(t)}{dt} = k_+c[1 - n(t)] - k_-n(t). \quad [2]$$

This equation describes the kinetics whereby the system comes to equilibrium. The free energy F associated with binding, which is given by the difference in the free energies of the unbound and bound states of the receptor, is related to the rate constant through detailed balance,

$$\frac{k_+c}{k_-} = \exp\left(\frac{F}{k_B T}\right). \quad [3]$$

If we imagine that thermal fluctuations can lead to small changes δk_+ and δk_- in the rate constants, we can linearize Eq. 2 to obtain

$$\frac{d\delta n}{dt} = -(k_+c + k_-)\delta n + c(1 - \bar{n})\delta k_+ - \bar{n}\delta k_-. \quad [4]$$

But from Eq. 3 we have

$$\frac{\delta k_+}{k_+} - \frac{\delta k_-}{k_-} = \frac{\delta F}{k_B T}. \quad [5]$$

Applying this constraint to Eq. 4, we find that the individual rate constant fluctuations cancel, and all that remains is the fluctuation in the thermodynamic binding energy δF ; the resulting equation can be written in the form

$$\frac{k_B T}{k_+c(1 - \bar{n})} \frac{d\delta n}{dt} + \frac{k_B T(k_+c + k_-)}{k_+c(1 - \bar{n})} \delta n = \delta F. \quad [6]$$

It is useful to note the analogy between this chemical kinetic problem and the Langevin equation (15) for the position, $X(t)$, of an overdamped Brownian particle bound by a Hookean spring. The spring generates a restoring force proportional to position, $-\kappa X$, and as the particle moves through the fluid it experiences a viscous drag with drag coefficient γ , so that the (Newtonian) equation of motion becomes

$$\gamma \frac{dX}{dt} + \kappa X = f(t), \quad [7]$$

where $f(t)$ is a fluctuating force. The dissipative and fluctuating parts of the force on the Brownian particle are related through the fluctuation–dissipation theorem

$$\langle f(t)f(t + \tau) \rangle = 2k_B T \gamma \delta(\tau), \quad [8]$$

where k_B is the Boltzmann constant and T is the temperature; angle brackets denote ensemble averages. Intuitively, this relation is a consequence of the fact that the fluctuating and the dissipative forces both arise because of collisions of the Brownian particle with the molecules of the fluid.

More generally, the linear response, $X(t)$, of a system from equilibrium due to the thermodynamically conjugate “force,” $F(t)$, defines the generalized susceptibility, $\alpha(t)$,

$$X(t) = \int_0^\infty \alpha(t')F(t - t')dt', \quad [9]$$

where we have taken $\langle X(t) \rangle = 0$ (Fig. 2). The generalized susceptibility depends on the properties of the system and completely characterizes its response to small external perturbations. Fourier transforming

$$\tilde{\alpha}(\omega) = \int_0^\infty \alpha(t)e^{i\omega t}dt, \quad [10]$$

the response to an external force near equilibrium becomes $\tilde{X}(\omega) = \tilde{\alpha}(\omega)\tilde{F}(\omega)$. In its general form, the fluctuation–dissipation theorem relates the imaginary part of the generalized susceptibility, $\tilde{\alpha}(\omega)$, which determines how much energy is dissipated by a system as heat due to an external force, to the power spectrum of the spontaneous fluctuations of the corresponding coordinate, X , for the closed system in thermal equilibrium

$$S_X(\omega) = \frac{2k_B T}{\omega} \text{Im}[\tilde{\alpha}(\omega)], \quad [11]$$

where $\text{Im}[\dots]$ refers to the imaginary part. In the present chemical system, the “coordinates” are the concentrations of the

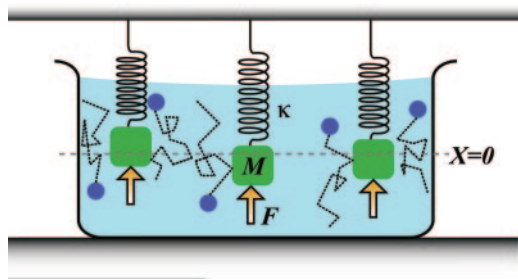


Fig. 2. For the mass-spring system immersed in a viscous fluid, measuring the linear response of the position, $X(t)$, to a known, small external force, $F(t)$, determines the generalized susceptibility. From this susceptibility, the fluctuation–dissipation theorem can be used to obtain the power spectrum of fluctuations in the closed system at equilibrium, as in Eq. 11. These fluctuations provide a lower bound on the accuracy of any measurement of the position. If measurements are carried out on N identical mass-spring systems, the expected error is reduced by a factor of $1/\sqrt{N}$. However, as N increases, this improvement ceases to hold, as neighboring mass-spring systems become physically close enough to experience correlated fluctuations from collisions with the particle bath, as shown. These correlations have been measured for two optically trapped colloidal particles (16).

interacting species, or equivalently the fractional occupancy of receptors, the phenomenological “equations of motion” are the chemical kinetic equations, and the thermodynamically conjugate “forces” are the free-energy differences among the species (17, 18) (Table 1).

Fourier transforming, from Eq. 6 we find the generalized susceptibility, $\tilde{\alpha}(\omega)$, describing the response of the coordinate n to its conjugate force F ,

$$\tilde{\alpha}(\omega) = \frac{\delta \bar{n}(\omega)}{\delta \tilde{F}(\omega)} = \frac{1}{k_B T} \frac{k_+ c(1 - \bar{n})}{-i\omega + (k_+ c + k_-)}. \quad [12]$$

The fluctuation–dissipation theorem relates this response function to the power spectrum of fluctuations in the occupancy, n ,

$$S_n(\omega) = \frac{2k_+ c(1 - \bar{n})}{\omega^2 + (k_+ c + k_-)^2}, \quad [13]$$

$$= \langle (\delta n)^2 \rangle \frac{2\tau_c}{1 + (\omega\tau_c)^2}, \quad [14]$$

where the total variance is

$$\langle (\delta n)^2 \rangle = \int_{-\infty}^{\infty} \frac{d\omega}{2\pi} S_n(\omega) = k_B T \left. \frac{\delta \bar{n}(\omega)}{\delta \tilde{F}(\omega)} \right|_{\omega=0} \quad [15]$$

$$= \frac{k_+ c(1 - \bar{n})}{k_+ c + k_-} = \bar{n}(1 - \bar{n}), \quad [16]$$

and the correlation time is given by $\tau_c = (k_+ c + k_-)^{-1}$. These are the usual results for switching in a Markovian way between two states; here it follows from the “macroscopic” kinetic

equations, as in Eq. 2, plus the fact that binding is an equilibrium process. Note that in using the fluctuation–dissipation theorem to arrive at this result, no assumptions are required about the underlying statistics of transitions between the bound and unbound states of the receptor. The Markovian nature of these transitions is reflected in the macroscopic chemical kinetic equations.

Coupled Binding and Diffusion. The same methods can be used in the more general case where the concentration itself has dynamics due to diffusion. Now we write

$$\frac{dn(t)}{dt} = k_+ c(\bar{x}_0, t)[1 - n(t)] - k_- n(t), \quad [17]$$

where the receptor is located at \bar{x}_0 , and

$$\frac{\partial c(\bar{x}, t)}{\partial t} = D \nabla^2 c(\bar{x}, t) - \delta(\bar{x} - \bar{x}_0) \frac{dn(t)}{dt}, \quad [18]$$

where the last term expresses the “injection” of one molecule at the point \bar{x}_0 as it unbinds from the receptor. Linearizing the equations as before, and solving Eq. 18 by transforming to spatial Fourier variables, we find the linear response function

$$\frac{\delta \bar{n}(\omega)}{\delta \tilde{F}(\omega)} = \frac{k_+ \bar{c}(1 - \bar{n})}{k_B T} \frac{1}{-i\omega[1 + \Sigma(\omega)] + (k_+ \bar{c} + k_-)}, \quad [19]$$

where \bar{c} is the mean concentration, and

$$\Sigma(\omega) = k_+(1 - \bar{n}) \int \frac{d^3 k}{(2\pi)^3} \frac{1}{-i\omega + Dk^2}. \quad [20]$$

We note that by obtaining the spatial Fourier transform of Eq. 18 over infinite volume, we are assuming the number of ligand molecules to be infinite at constant concentration. Hence, we are considering only the regime where the number of ligand molecules exceeds the number of receptors. The “self-energy” $\Sigma(\omega)$ is ultraviolet divergent, which can be traced to the delta function in Eq. 18; we have assumed that the receptor is infinitely small. A more realistic treatment would give the receptor a finite size, which is equivalent to cutting off the k integrals at some (large) $\Lambda \sim \pi/a$, with a the linear dimension of the receptor.

If we imagine mechanisms that read out the receptor occupancy and average over a time τ long compared with the correlation time τ_c of the noise, then the relevant quantity is the low frequency limit of the noise spectrum. Hence, we are interested in

$$\Sigma(\omega \ll D/a^2) \approx \Sigma(0) = \frac{k_+(1 - \bar{n})}{2\pi Da}, \quad [21]$$

and

Table 1. Linear response in mechanical and chemical systems

Physical quantity	Mass-spring system	Chemical system
Coordinate	Displacement	Receptor occupancy, $\delta n = n - \bar{n}$
Conjugate force	f	Free energy change $\delta F = k_B T (\delta k_+ / k_+ - \delta k_- / k_-)$
Spring constant	κ	$k_B T / [\bar{n}(1 - \bar{n})]$
Damping constant	γ	$k_B T / (k_- \bar{n})$

The response of a chemical system near equilibrium is directly analogous to that of the familiar mass-spring system in a viscous fluid, in the limit that the inertial term, $M\ddot{X}$, can be neglected (valid for $\kappa/M \ll \gamma^2/4$).

$$\frac{\delta\bar{n}(\omega)}{\delta\bar{F}(\omega)} = \frac{k_+\bar{c}(1-\bar{n})}{k_{\text{BT}}} [-i\omega(1+\Sigma(0)) + (k_+\bar{c} + k_-)]^{-1}. \quad [22]$$

Applying the fluctuation–dissipation theorem once again, we find

$$S_n(\omega) \approx \frac{2k_+\bar{c}(1-\bar{n}) [1 + \Sigma(0)]}{\omega^2 [1 + \Sigma(0)]^2 + (k_+\bar{c} + k_-)^2}. \quad [23]$$

The total variance in occupancy is unchanged because it is an equilibrium property of the system. Coupling to concentration fluctuations does serve to renormalize the correlation time of the noise, $\tau_c \rightarrow \tau_c [1 + \Sigma(0)]$. The new τ_c can be written as

$$\tau_c = \frac{1-\bar{n}}{k_-} + \frac{\bar{n}(1-\bar{n})}{2\pi D a \bar{c}}, \quad [24]$$

so the second term is a lower bound on τ_c , independent of the kinetic parameters k_{\pm} ,

$$\tau_c > \frac{\bar{n}(1-\bar{n})}{2\pi D a \bar{c}}. \quad [25]$$

Again, the relevant quantity is the low-frequency limit of the noise spectrum,

$$S_n(\omega=0) = \frac{2\bar{n}(1-\bar{n})}{k_+\bar{c} + k_-} + \frac{[\bar{n}(1-\bar{n})]^2}{\pi D a \bar{c}}. \quad [26]$$

If we average for a time τ , then the root-mean-square error in our estimate of n will be

$$\delta n_{\text{rms}} = \sqrt{S_n(0) \cdot \frac{1}{\tau}}, \quad [27]$$

and we see that this noise level has a minimum value independent of the kinetic parameters k_{\pm} ,

$$\delta n_{\text{rms}} > \frac{\bar{n}(1-\bar{n})}{\sqrt{\pi D a \bar{c}} \tau}. \quad [28]$$

To relate these results back to the discussion by Berg and Purcell (6), we note that an overall change in concentration is equivalent to a change in F by an amount equal to the change in chemical potential, so that $\Delta c/\bar{c} \equiv \Delta F/k_{\text{BT}}$. This equivalence means that there is an effective spectral density of noise in measuring c

$$S_c^{\text{eff}}(\omega) = \left(\frac{\bar{c}}{k_{\text{BT}}}\right)^2 S_F(\omega), \quad [29]$$

where the “noise force” spectrum $S_F(\omega)$ is given by the fluctuation–dissipation theorem as

$$S_F(\omega) = \left| \frac{\delta\bar{n}(\omega)}{\delta\bar{F}(\omega)} \right|^{-2} S_n(\omega) = -\frac{2k_{\text{BT}}}{\omega} \text{Im} \left[\frac{\delta\bar{F}(\omega)}{\delta\bar{n}(\omega)} \right]. \quad [30]$$

In the present case, we find that

$$S_c^{\text{eff}}(\omega) = \frac{2\bar{c}^2}{k_+\bar{c}(1-\bar{n})} + \frac{\bar{c}}{\pi D a}. \quad [31]$$

As before, the accuracy of a measurement that integrates for a time τ is set by

$$\delta c_{\text{rms}} = \sqrt{S_c^{\text{eff}}(0) \cdot \frac{1}{\tau}}, \quad [32]$$

and we find again a lower bound that is determined only by the physics of diffusion,

$$\frac{\delta c_{\text{rms}}}{\bar{c}} > \frac{1}{\sqrt{\pi D a \bar{c}} \tau}. \quad [33]$$

Note that this result is (up to a factor of $\sqrt{\pi}$) exactly the Berg–Purcell formula in Eq. 1.

Binding to Multiple, Noninteracting Receptors. To complete the derivation of Berg and Purcell’s original results (6), we consider a collection of m receptor sites at positions \vec{x}_μ , where $\mu = 1, 2, \dots, m$:

$$\frac{dn_\mu(t)}{dt} = k_+c(\vec{x}_\mu, t)[1 - n_\mu(t)] - k_-n_\mu(t), \quad [34]$$

$$\frac{\partial c(\vec{x}, t)}{\partial t} = D\nabla^2 c(\vec{x}, t) - \sum_{i=1}^m \delta(\vec{x} - \vec{x}_\mu) \frac{dn_\mu(t)}{dt}. \quad [35]$$

We can solve Eq. 35 by going to a spatial Fourier representation as before, and we find

$$\delta c(\vec{x}_\nu, \omega) = \frac{i\omega\Lambda}{2\pi^2 D} \delta\bar{n}_\nu(\omega) + \frac{i\omega}{2\pi^2} \sum_{\mu \neq \nu}^m \frac{\delta\bar{n}_\mu(\omega)}{|\vec{x}_\nu - \vec{x}_\mu|} \int_0^\infty \frac{k \sin(k|\vec{x}_\nu - \vec{x}_\mu|)}{-i\omega + Dk^2} dk, \quad [36]$$

where Λ is the cut-off wave number; as before, the cut-off arises to regulate the delta function in Eq. 35 and is related to the size of the individual receptor. In the limit $|\vec{x}_\nu - \vec{x}_\mu| (\omega/D)^{1/2} \ll 1$, for $\nu \neq \mu$ with $\nu, \mu = 1, \dots, m$, we have

$$\delta c(\vec{x}_\nu, \omega) = \frac{i\omega\Lambda}{2\pi^2 D} \delta\bar{n}_\nu(\omega) + \frac{i\omega}{4\pi D} \sum_{\mu \neq \nu}^m \frac{\delta\bar{n}_\mu(\omega)}{|\vec{x}_\nu - \vec{x}_\mu|}. \quad [37]$$

Combining this equation with the Fourier transform of Eq. 34 and summing to find the total occupancy $\delta\bar{N}(\omega) = \sum_{\mu=1}^m \delta n_\mu(\omega)$ of the receptor cluster, we obtain

$$-i\omega\delta\bar{N} = - \left[(k_+\bar{c} + k_-) - \frac{i\omega\Lambda k_+(1-\bar{n})}{2\pi^2 D} \right] \delta\bar{N} + \frac{i\omega k_+(1-\bar{n})}{4\pi D} \sum_{\nu=1}^m \sum_{\mu \neq \nu} \delta\bar{n}_\mu \frac{1}{|\vec{x}_\mu - \vec{x}_\nu|} + mk_+\bar{c}(1-\bar{n}) \left(\frac{\delta\bar{F}}{k_{\text{BT}}} \right). \quad [38]$$

In cluster geometries such that the innermost sum is independent of \vec{x}_ν , we can write the sum as

$$\sum_{\nu=1}^m \sum_{\mu \neq \nu} \delta\bar{n}_\mu \frac{1}{|\vec{x}_\mu - \vec{x}_\nu|} = \phi(m) \cdot \delta\bar{N}, \quad [39]$$

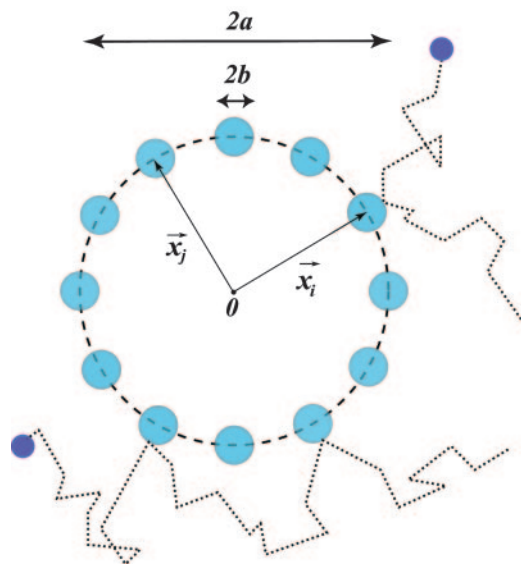


Fig. 3. Schematic representation of a cluster of m receptors of size b , distributed uniformly on a ring of size a . For $a \gg b$, the relative accuracy in measurement of the substrate concentration improves as $1/\sqrt{m}$ until $mb \sim a$, at which point the binding/unbinding events of nearby receptors are no longer independent.

where

$$\phi(m) = \sum_{\mu=2}^m \frac{1}{|\bar{x}_{\mu} - \bar{x}_1|}, \quad [40]$$

and this simplification allows us to solve Eq. 38 directly to find the response of $\delta\bar{N}$ to the force $\delta\bar{F}$. Then, as before, we use the fluctuation–dissipation theorem to find the spectrum of $\delta\bar{F}$ and convert that to an equivalent concentration error as in Eq. 31. The result is

$$\left(\frac{\delta c_{\text{rms}}}{\bar{c}}\right)^2 \approx \frac{2}{mk_+ \bar{c}(1-\bar{n})\tau} + \frac{1}{\pi D \bar{c} \tau} \left(\frac{\Lambda}{m} + \frac{\phi(m)}{2m}\right). \quad [41]$$

We note that whereas the first term is positive and depends on the details of the chemical kinetics of ligand–receptor binding, the second term defines a lower bound on the measurement accuracy of the ligand concentration by the receptor cluster that depends only on the physics of diffusion. As an example, for receptors of radius b uniformly distributed around a ring of radius $a > b$ (Fig. 3), this lower bound is

$$\frac{\delta c_{\text{rms}}}{\bar{c}} > \frac{1}{\sqrt{\pi D \bar{c} \tau}} \left(\frac{1}{mb} + \frac{g_0}{2a}\right)^{1/2}, \quad [42]$$

where for $m \gg 1$, we have used $\phi(m) \approx mg_0/a$; g_0 is a geometric factor of order unity for typical cluster geometries and receptor distributions (W.B. and S.S., unpublished results). With an increasing number of receptors, m , the accuracy in measuring the substrate concentration ultimately is limited by the linear size of the cluster. Extension of this result to cooperatively interacting receptors is treated separately elsewhere (W.B. and S.S., unpublished results); remarkably, we find that this lower bound in measurement accuracy persists, independent of the details of cooperative interactions among the cluster subunits.

Summary. For both a single receptor and an array of receptors, we find that the sensitivity of signaling is limited, and this limit can be described as an effective noise in concentration, δc_{rms} , determined by

$$\left(\frac{\delta c_{\text{rms}}}{\bar{c}}\right)^2 = F(\{k_j\}, \bar{c}, m) + \frac{1}{\pi D \bar{c} \tau \ell}. \quad [43]$$

In this summary, $F(\{k_j\}, \bar{c}, m)$ depends on the details of the kinetic interactions between the signaling molecules and its receptor through the kinetic parameters, $\{k_j\}$, the substrate concentration, \bar{c} , and the number of receptors, m . The important point is that this term is positive, so that even if we do not know the details of the ligand–receptor chemical kinetics, we know that the noise level can never be smaller than that set by the second term. In the second term ℓ is an effective size for the receptor or the receptor array; then, to within a factor, this term is exactly that written down by Berg and Purcell (Eq. 1) for a perfect concentration measuring device. Thus, although the original Berg and Purcell (6) arguments made use of very specific assumptions, we see that in the general case the details of the chemical signaling only add to the noise level.

Comparison with Experiment

Here, we consider two experimentally well-characterized examples to demonstrate how the theoretical limits on receptor occupancy noise and the resulting precision of concentration measurements compare with the performance of real cellular signaling systems.

Regulation of Gene Expression in Bacteria. Expression of genes is controlled in part by the occupancy of specific sites in the promoter regions adjacent to the sequences of DNA that code for protein (19). Reversing the usual picture of changes in transcription factor concentration as driving changes in gene expression, we can view gene expression as a sensor for the concentration of the transcription factor proteins that bind to the promoter site. In a bacterium like *E. coli*, transcription factors are present in $N_{\text{TF}} \sim 100$ copies in a cell of volume of $\sim 1 \mu\text{m}^3$ (20); presumably, the concentration of free transcription factor molecules is smaller than N_{TF}/V . Diffusion constants for small proteins in the *E. coli* cytoplasm are $D \sim 3 \mu\text{m}^2/\text{s}$ (21); promoter sites have a linear dimension $a \sim 3 \text{ nm}$, and putting these factors together, we find the crucial combination of parameters $\pi D a \bar{c} \approx 3 \text{ s}^{-1}$.

If the transcription factor is a repressor then gene expression levels are determined by $1 - n$, whereas if it is an activator then expression is related to n . Because $\delta n_{\text{rms}} \propto \bar{n}(1 - \bar{n})$ (Eq. 28), fractional fluctuations in either $A = n$ or $A = 1 - n$ are determined by

$$\frac{\delta A}{\bar{A}} = (1 - \bar{A}) \frac{1}{\sqrt{\pi D a \bar{c} \tau}}. \quad [44]$$

The minimum fluctuations in expression level thus are given by

$$\frac{\delta A}{\bar{A}} > (0.1) \cdot (1 - \bar{A}) \cdot \left(\frac{100}{N_{\text{TF}}}\right)^{1/2} \cdot \left(\frac{30 \text{ s}}{\tau}\right)^{1/2}. \quad [45]$$

Recent experiments (22) indicate that *E. coli* achieves $\approx 10\%$ precision in control of gene expression at small values of \bar{A} . For this performance to be consistent with the physical limits, the transcription machinery must therefore integrate the promoter site occupancy for times of the order of 1 min, even assuming that the translation from occupancy to expression level itself is noiseless. This integration can be provided by the lifetime of the mRNA transcripts themselves, which is $\sim 3 \text{ min}$ in prokaryotes (23). Recent theoretical work, motivated by ref. 22, has ad-

