



HAWAII



PRINCETON



PENNSYLVANIA STATE UNIVERSITY



AFOSR MURI Update June 2005-Jan 2006 :

Renewable Bio-solar Hydrogen Production from Robust Oxygenic Phototrophs

BioSolarH₂ → Team

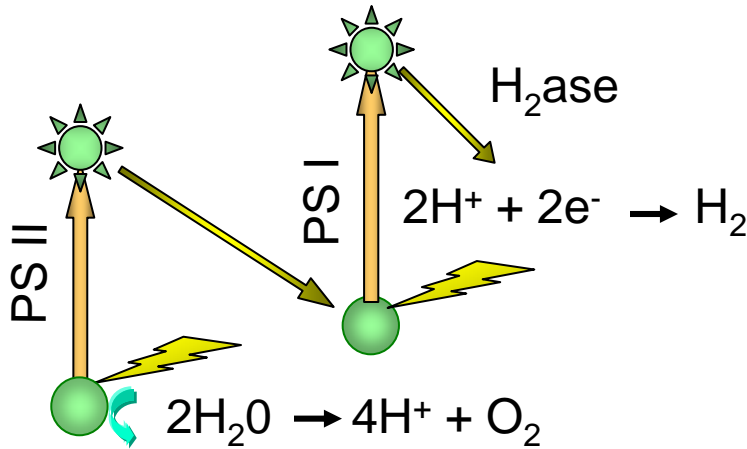
G. Charles Dismukes	PU	photosynthesis/engineering
Edward I. Stiefel	PU	enzymology/inorganic
Donald A. Bryant	PSU	genetics/mol biology cyanos
Matthew Posewitz	CSM	genetics/mol biology algae
Eric Hegg	UU	enzymology/inorganic
Robert Bidigare	UH	microbial physiology/ecology



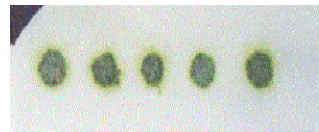
-BioSolarH₂→



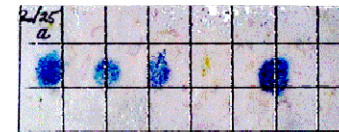
Goal 1: High Throughput Screening for BioSolar H₂ Production



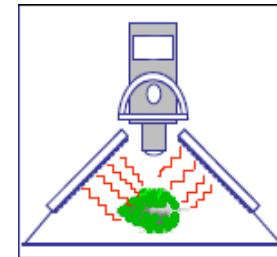
Colonies on agar plates



Chemochromic H₂ sensor



Kinetic Fluorescence CCD Camera



Goal: screen libraries of phototrophs in search of most active H₂ producers using multiple sensors for H₂, photosynthetic capacity and O₂ consumption

Payoff to Air Force

- development of robust screening protocol to identify unique H₂ producing organisms
- development of clean renewable hydrogen fuel



-BioSolarH₂ →

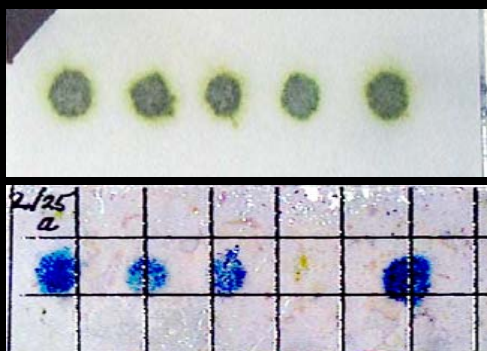


Goal 2: Design/Fabricate Powerful Instrumentation

Current Tools for:

- high throughput screening of gaseous H₂ production
- Light induced electron transfer rates in/out of PSII: H₂O → O₂, Q_A⁻ → Q_A
- Intracellular redox status PSII: PQ/PQH₂
- Nanomolar sensitivity for dissolved O₂ and H₂ concentrations

Colonies on agar



Fast repetition rate fluorimeter
PSII Quantum Efficiency



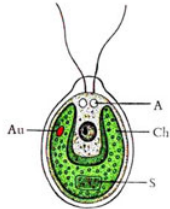
Dissolved H₂ & O₂
LED + Clark cells



NREL H₂ gas sensor



-BioSolarH₂ →



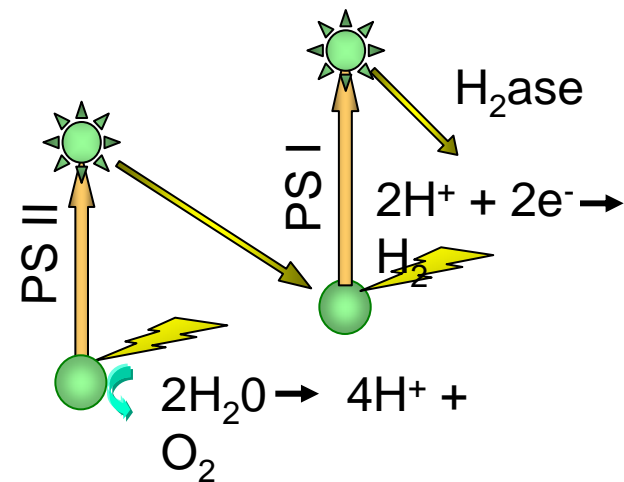
Goal 3: Elucidate Pathways for e⁻ & H⁺ fluxes in Microalgae: Light-Induced H₂ Production in *Chlamydomonas reinhardtii*

Mechanistic Pathway in WT strain cc124

- Two pools of photo-electron acceptors in PSI identified as precursors to photo-H₂
- Established conditions for absence of competing pathways to photo-H₂

Genetically Engineered Strains

- Expression of a single set of H₂ase assembly genes has been demonstrated to be sufficient to assemble a diverse set of [FeFe]-hydrogenase structural enzymes from foreign *HydA* genes taken from clostridial bacterial strains.

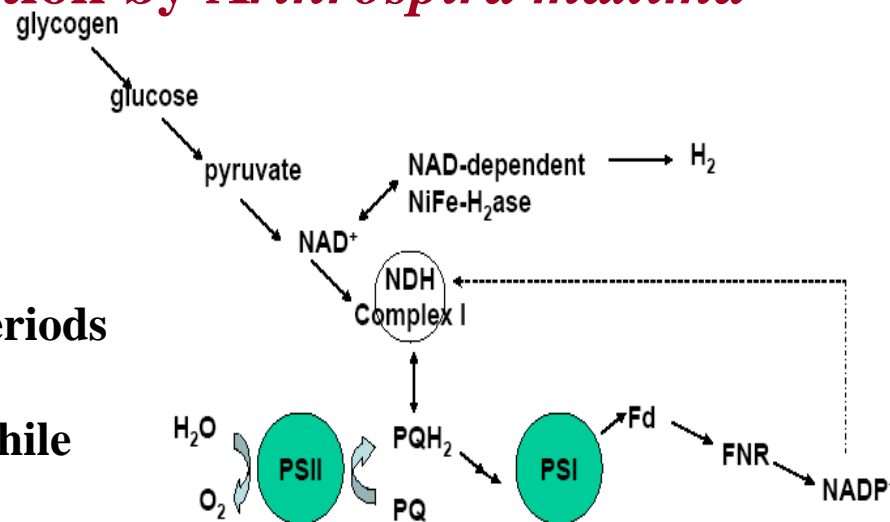




-BioSolarH₂ →



Goal 3: Elucidate Pathways for e⁻ & H⁺ Fluxes in Cyanobacteria: Dark Fermentative H₂ production by *Arthrospira maxima*



- Robust photoautotrophic growth over long periods
- High carbonate-requiring hypersaline alkalophile
- Optimal temporal separation of H₂ and O₂
- High rates of fermentative H₂ production optimized by environment conditions (T, pH) and micro-nutrient optimization (Ni, Fe, trace metals): 5.5 ml H₂ (liter culture)⁻¹ hour⁻¹

Two stage indirect pathway to generate H₂ via a O₂-tolerant NiFe-H₂ase:

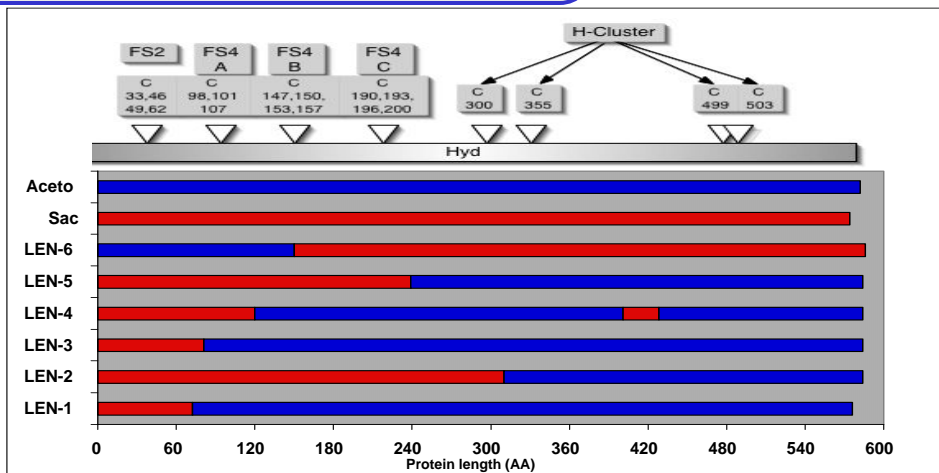
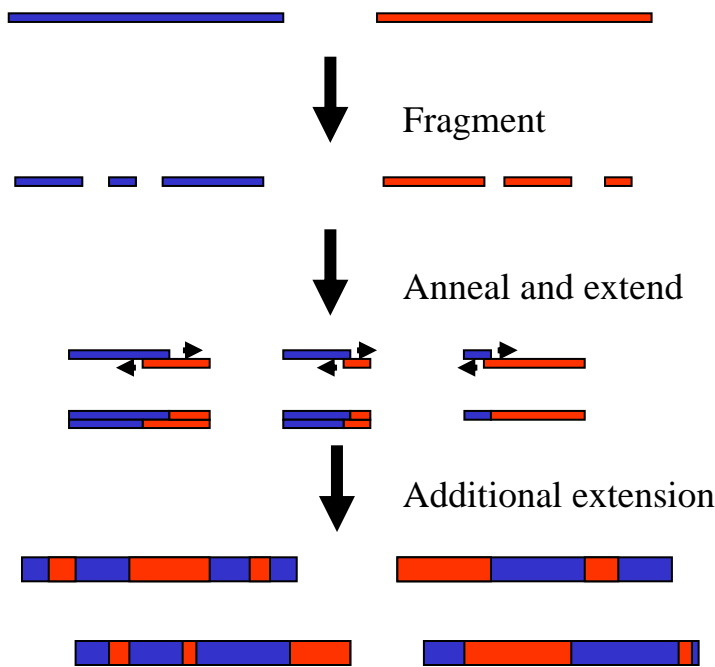
- 1) classical photosynthetic generation of glycogen,
- 2) fermentation of glycogen



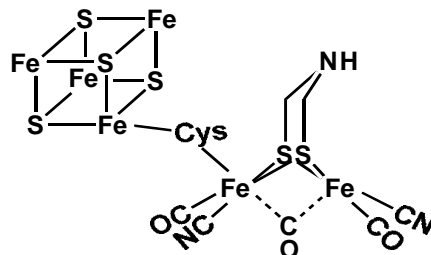
Goal 4: Gene shuffling to rapidly increase diversity of [FeFe]-hydrogenases

Goal: Use recombinant DNA technology to rapidly generate large libraries of novel [FeFe]-hydrogenases. Select robust enzymes for application in BioSolar H₂-production applications

[FeFe]-hydrogenase (A) [FeFe]-hydrogenase (B)



Examples of shuffled products with activity



FeFe-hydrogenase catalytic H-cluster

Payoff to Air Force

- Rapid generation of millions of novel [FeFe]-hydrogenases in less than a week
- Efficiently generates enzyme libraries to be screened for more robust properties
 - Established biotechnology for improving enzyme function